

# Automating high-throughput methylomes

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# Introduction



*Name:* Lee M Butcher

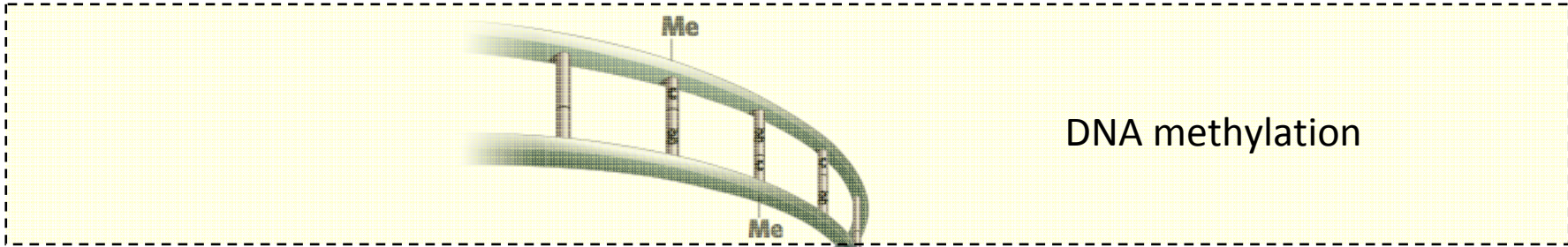
*Position:* Postdoc

*Team:* Medical Genomics

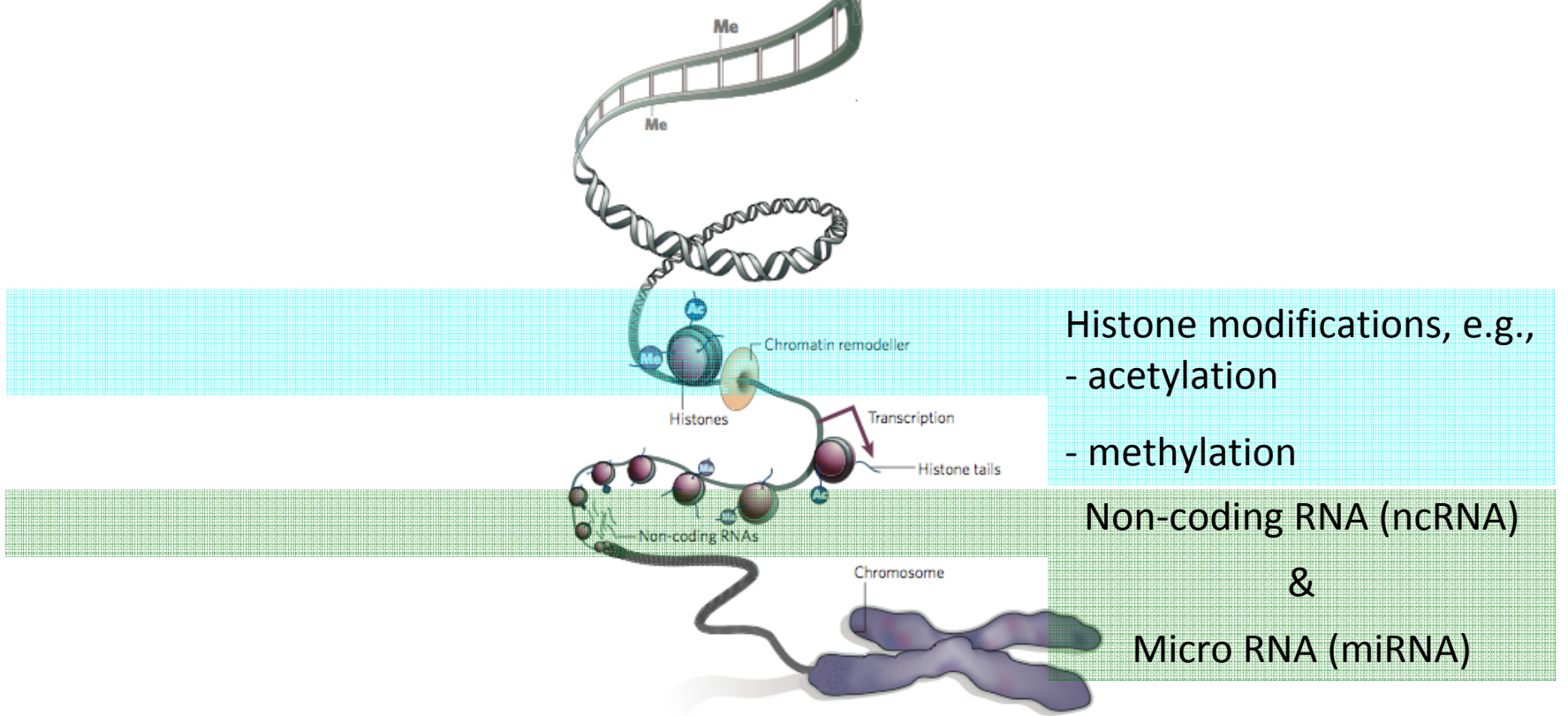
*Supervisor:* Stephan Beck

*Lab interests:* High-throughput technology  
Epigenetics/epigenomics  
Human complex disease/traits  
Mouse development

# Epigenetics



DNA methylation



Histone modifications, e.g.,  
- acetylation

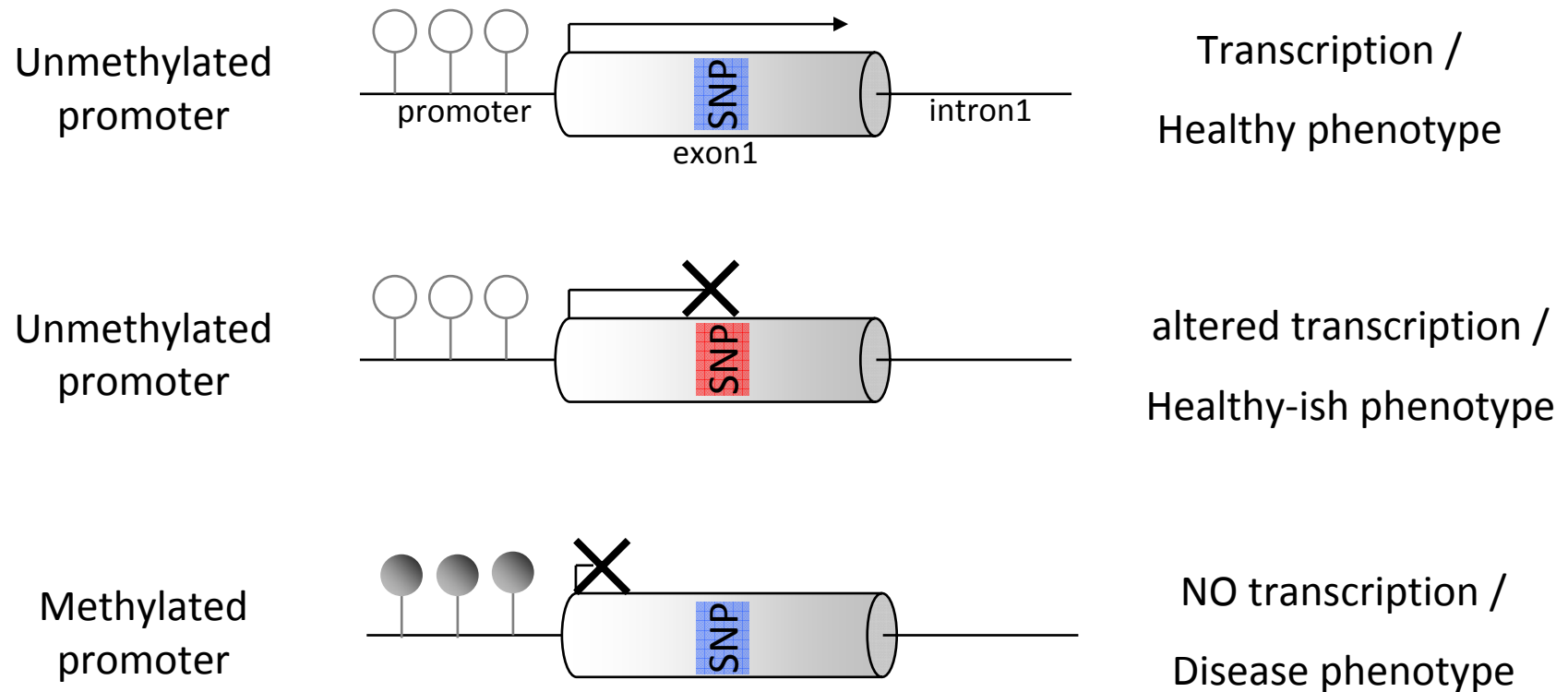
- methylation

Non-coding RNA (ncRNA)

&

Micro RNA (miRNA)

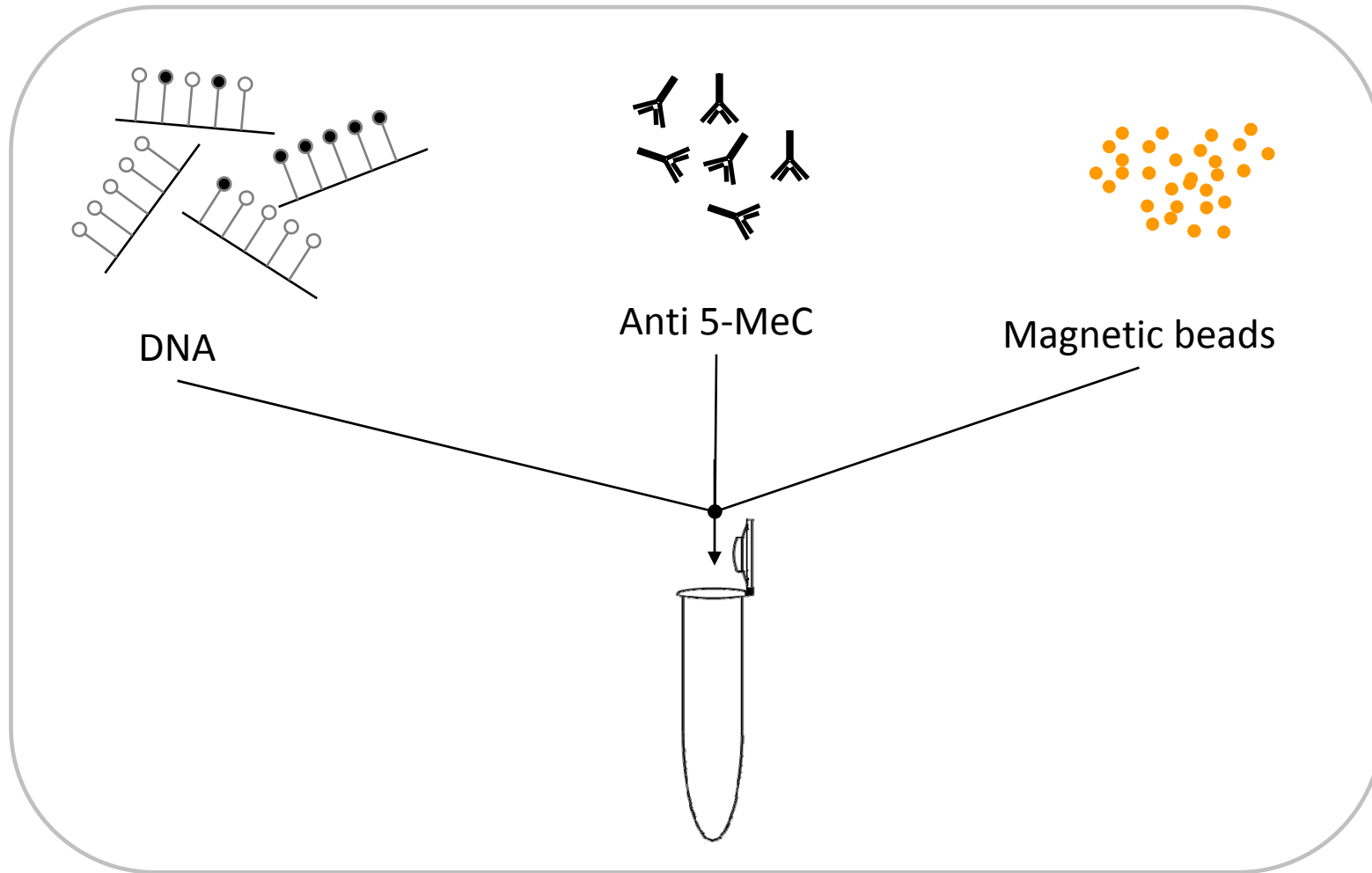
# Genomics and epigenomics



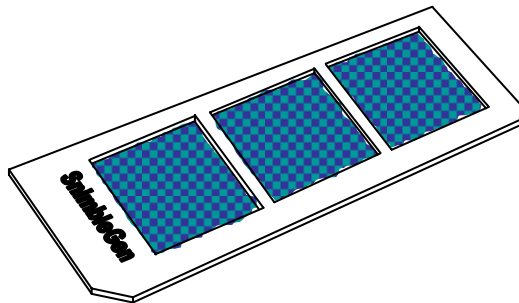
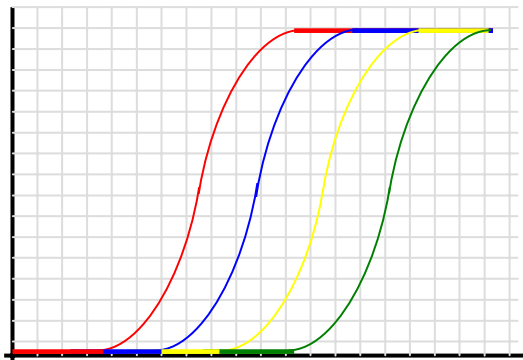
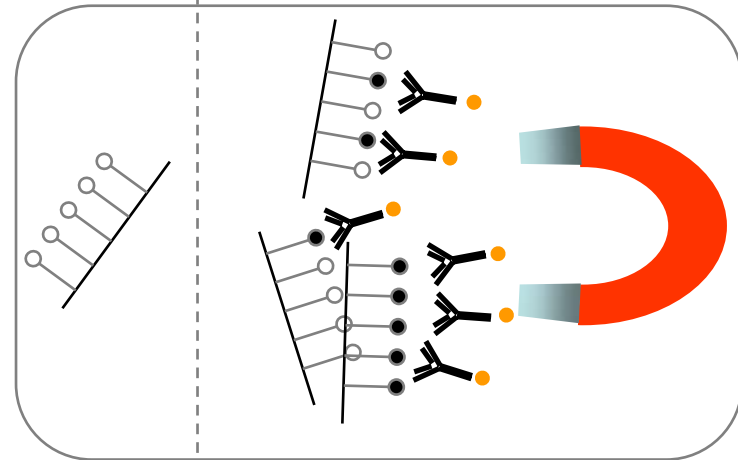
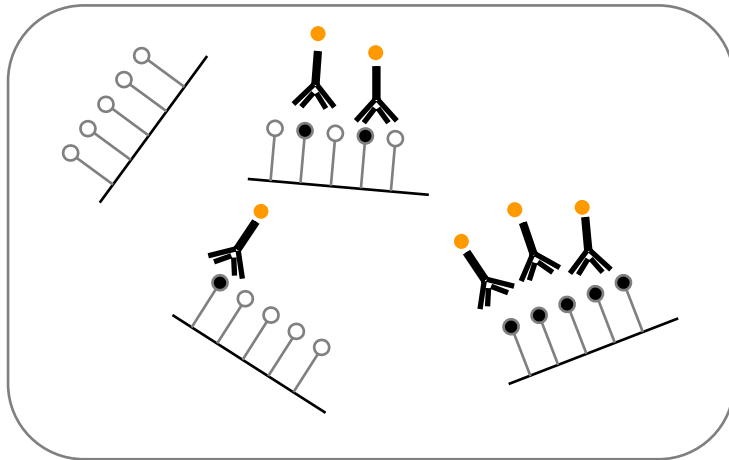
# Challenges for epigenomic research

- Complex disease is influenced by multiple genes
  - Disease loci scattered at [largely] unknown genomic positions
  - Need an epigenetic assay that is:
    - Accurate
    - Affordable
    - High-throughput
    - Whole genome
- Assay of choice:
  - Methylated DNA immunoprecipitation (MeDIP; Weber et al. [2005] NatGen)
  - Decent balance between:
    - Throughput
    - Genomic coverage
    - Cost

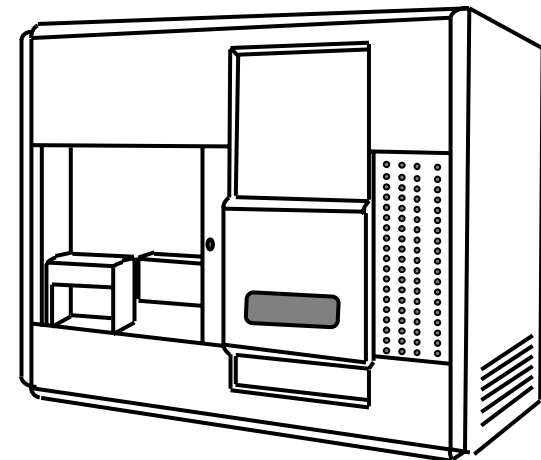
# MeDIP assay – how it works



# MeDIP assay – how it works



{MeDIP-Chip}

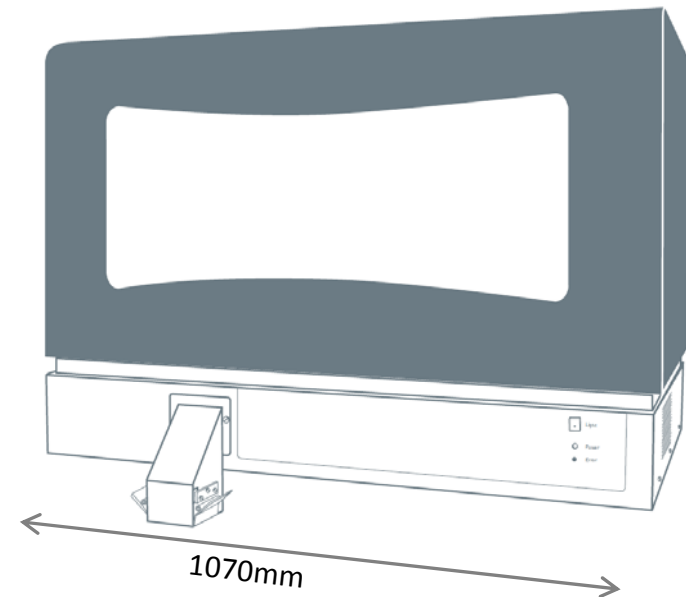


{MeDIP-Seq}

# AutoMeDIP



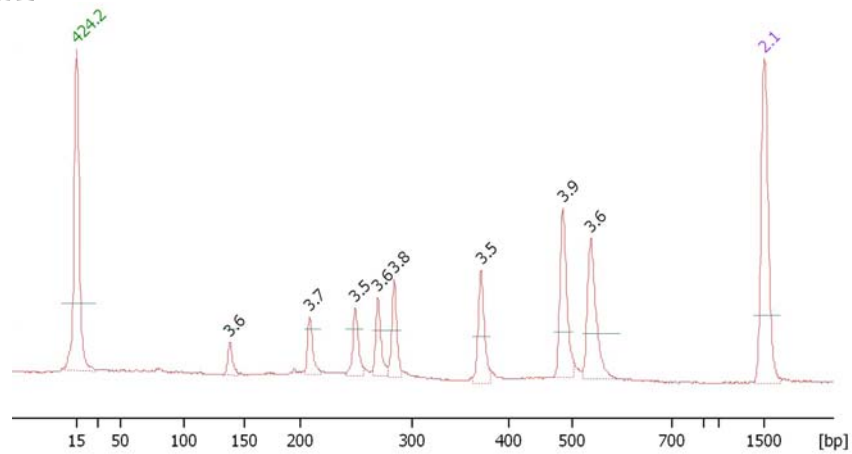
- Diagenode
  - Automates up to 16 samples per run
    - Up to 24 samples every 24 hours (5 hour IP)
  - Hands on time saving = 75%
- Performance measured using:
  - *In vitro* methylated spike
  - qPCR





# Spike-in made from $\lambda$ -DNA

**A**



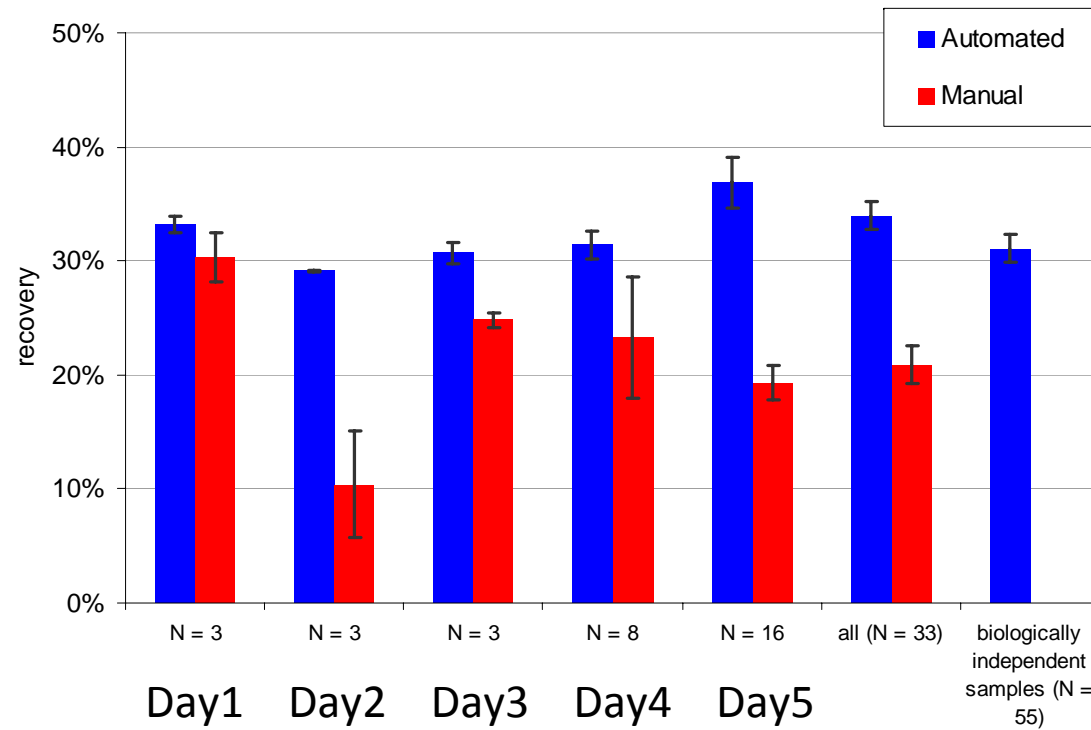
**B**

Size (bp)	No. CpGs	CpG density	Conc. (ng/ $\mu$ l)	Molarity (nM/ltr)
138	2	1.4%	0.33	3.6
197	1	0.5%	0.51	3.7
226	3	1.3%	0.58	3.5
270	5	1.9%	0.64	3.6
284	20	7.0%	0.7	3.8
360	10	2.8%	0.87	3.5
462	15	3.2%	1.25	3.9
400	20	5.0%	1.29	3.6

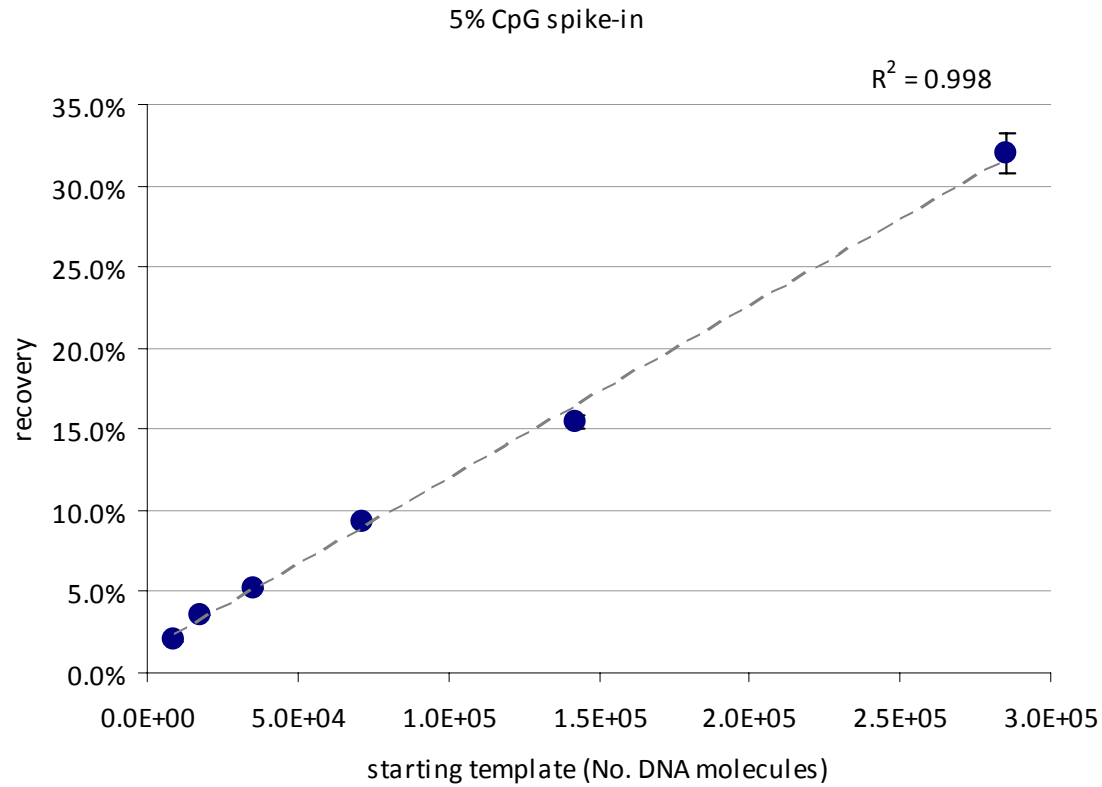
# Results



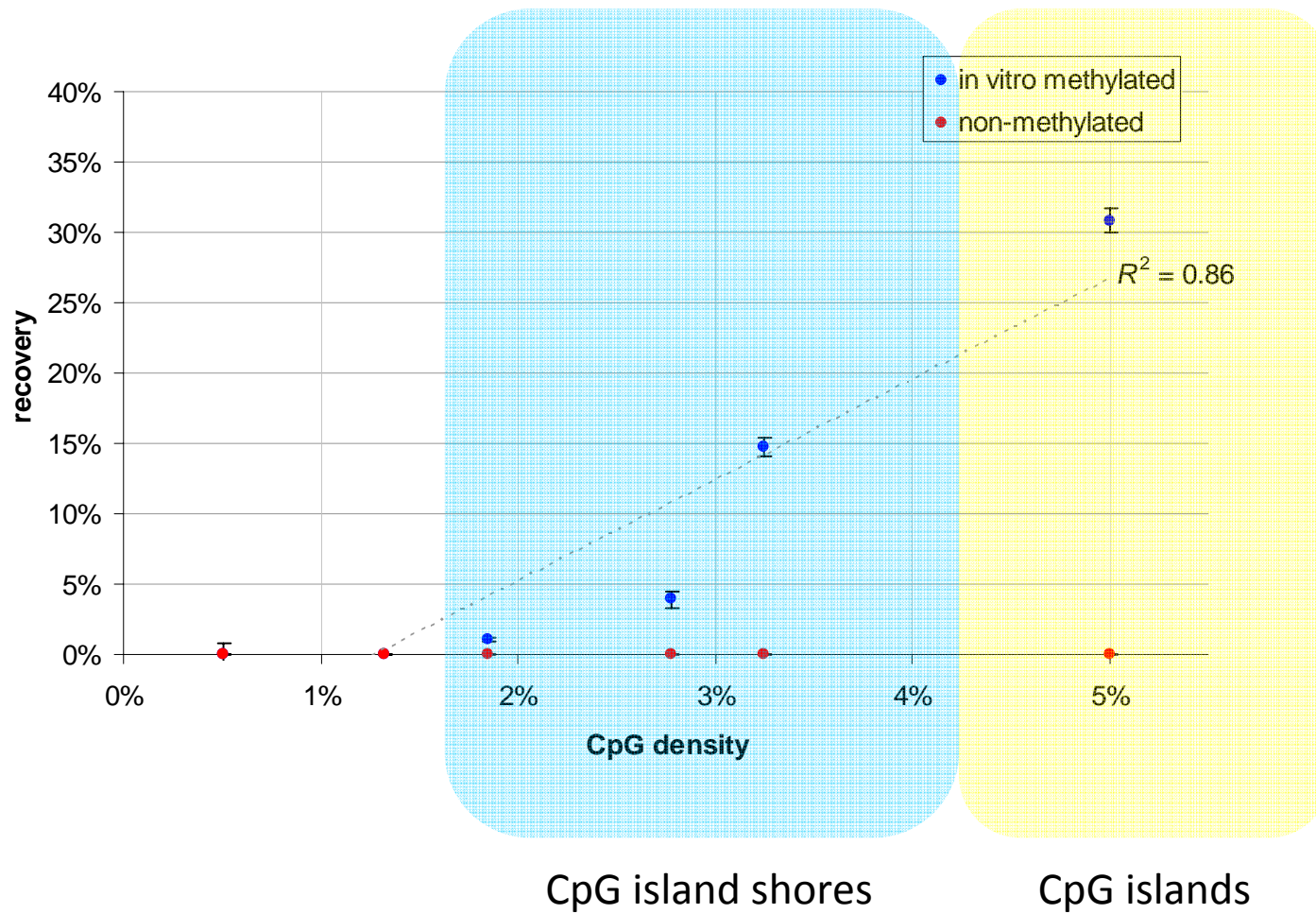
# Results – reproducibility



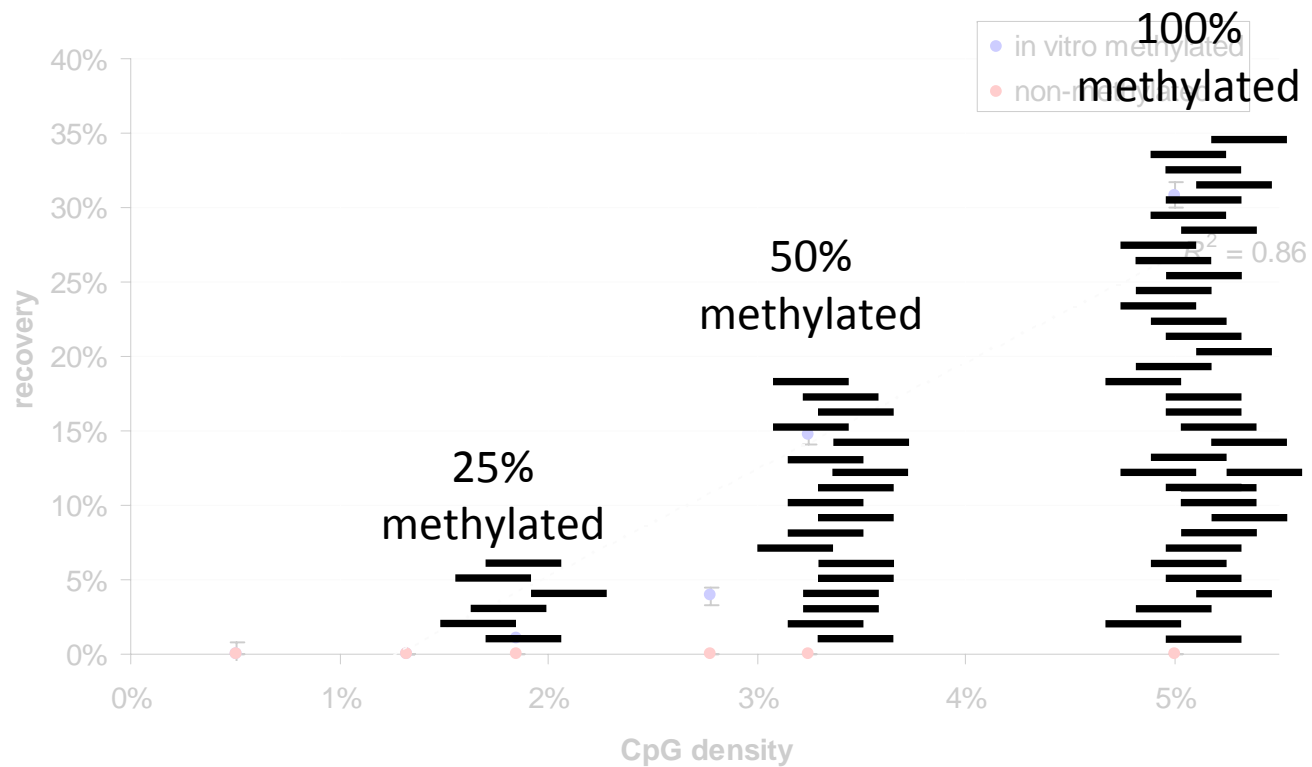
# Results – sensitivity



# Results – specificity



# Results – implications on alignment



# Bioinformatic challenges...



- Enrichment bias means absolute methylation levels are difficult to quantitate
- Fortunately, help is at hand:

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A Bayesian deconvolution strategy for immunoprecipitation-based DNA methylome analysis

Thomas A Down<sup>1,8</sup>, Vardhman K Rakyan<sup>2,8</sup>, Daniel J Turner<sup>3</sup>, Paul Flicek<sup>4</sup>, Heng Li<sup>3</sup>, Eugene Kulesha<sup>4</sup>, Stefan Gräf<sup>4</sup>, Nathan Johnson<sup>4</sup>, Javier Herrero<sup>4</sup>, Eleni M Tomazou<sup>3</sup>, Natalie P Thorne<sup>5</sup>, Liselotte Bäckdahl<sup>6</sup>, Marlis Herberth<sup>7</sup>, Kevin L Howe<sup>5</sup>, David K Jackson<sup>3</sup>, Marcos M Miretti<sup>3</sup>, John C Marioni<sup>5</sup>, Ewan Birney<sup>4</sup>, Tim J P Hubbard<sup>3</sup>, Richard Durbin<sup>3</sup>, Simon Tavaré<sup>5</sup> & Stephan Beck<sup>6</sup>

- Unfortunately...

$$f(A/m) = \prod_p G \left( A_p / A_{base} + r \sum_c C_{cp} m_c, \nu^{-1} \right)$$

# MeDIP and non-CG methylation

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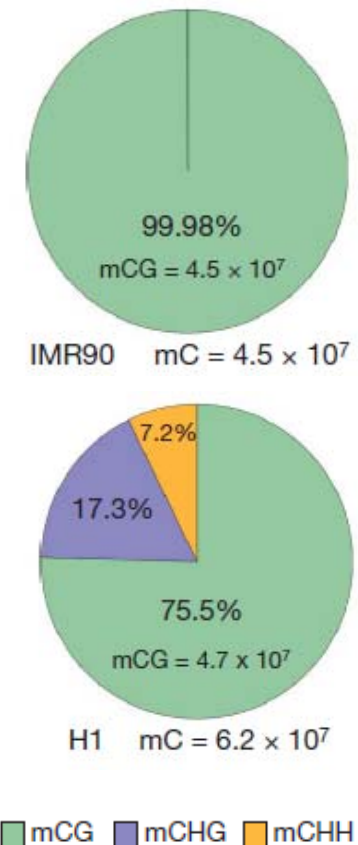
nature

ARTICLES

## Human DNA methylomes at base resolution show widespread epigenomic differences

Ryan Lister<sup>1\*</sup>, Mattia Pelizzola<sup>1\*</sup>, Robert H. Downen<sup>1</sup>, R. David Hawkins<sup>2</sup>, Gary Hon<sup>2</sup>, Julian Tonti-Filippini<sup>4</sup>, Joseph R. Nery<sup>1</sup>, Leonard Lee<sup>2</sup>, Zhen Ye<sup>2</sup>, Que-Minh Ngo<sup>2</sup>, Lee Edsall<sup>2</sup>, Jessica Antosiewicz-Bourget<sup>5,6</sup>, Ron Stewart<sup>5,6</sup>, Victor Ruotti<sup>5,6</sup>, A. Harvey Millar<sup>4</sup>, James A. Thomson<sup>5,6,7,8</sup>, Bing Ren<sup>2,3</sup> & Joseph R. Ecker<sup>1</sup>

DNA cytosine methylation is a central epigenetic modification that has essential roles in cellular processes including genome regulation, development and disease. Here we present the first genome-wide, single-base-resolution maps of methylated cytosines in a mammalian genome, from both human embryonic stem cells and fetal fibroblasts, along with comparative analysis of messenger RNA and small RNA components of the transcriptome, several histone modifications, and sites of DNA–protein interaction for several key regulatory factors. Widespread differences were identified in the composition and patterning of cytosine methylation between the two genomes. **Nearly one-quarter of all methylation identified in embryonic stem cells was in a non-CG context, suggesting that embryonic stem cells may use different methylation mechanisms to affect gene regulation.** Methylation in non-CG contexts showed enrichment in gene bodies and depletion in protein binding sites and enhancers. Non-CG methylation disappeared upon induced differentiation of the embryonic stem cells, and was restored in induced pluripotent stem cells. We identified hundreds of differentially methylated regions proximal to genes involved in pluripotency and differentiation, and widespread reduced methylation levels in fibroblasts associated with lower transcriptional activity. These reference epigenomes provide a foundation for future studies exploring this key epigenetic modification in human disease and development.





# MeDIP and non-CG methylation

*The CG context*



5' GCTTCTCTGGAGTGCCACAGGTTTGATGACAAAAATTAGCCCAAGAAGACAAAAATCACCTTGCCCTAATGCT 3'  
3' CGAAGAGACCTCACCCGTGCCAAACTACTGTTTTTTAATCCCGTTCTTCTGTTTTTAGTGGAACCCGATTCCGA 5'



# MeDIP and non-CG methylation

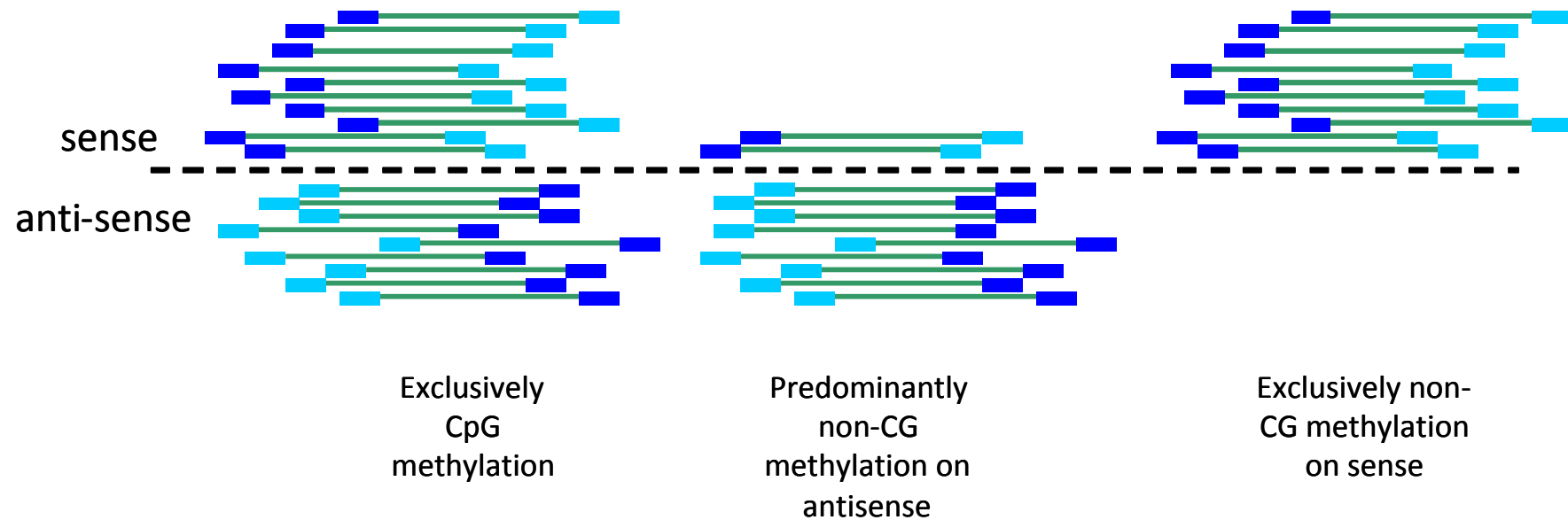
*The non-CG context*



5' GCTTCTCTGGAGTGCGACAGGTTTGATGACAAAATTAGCGCAAGAAGACAAAATCACCTGCGCTAATGCT 3'  
3' CGAAGAGACCTCACGCTGTCCAACTACTGTTTTTTAATCGCGTTCTTCTGTTTTTTAGTGGAACGCGATTCCGA 5'

# MeDIP and non-CG methylation

- Non-CG methylation will result in strand bias during sequencing



# Applications of MeDIP



- We've collaborated on:
  - Two “large-scale” projects ( $N=37$ ,  $N=60$ )
  - Four smaller-scale projects ( $N=3$ ,  $N=6$ ,  $N=7$ ,  $N=12$ )
- Allocate 1 lane of flow cell (Illumina GAIIx)
  - Generate 40M reads
  - Average genome-wide CpG depth  $\approx 1.2$ - $1.5x$
  - Average CpG depth covered by reads  $\approx 3.3$ - $4.4x$
  - No. CpGs covered by reads  $\approx 7.1M$ - $10.9M$  (31.8% - 48.8%)

# A cancer methylome

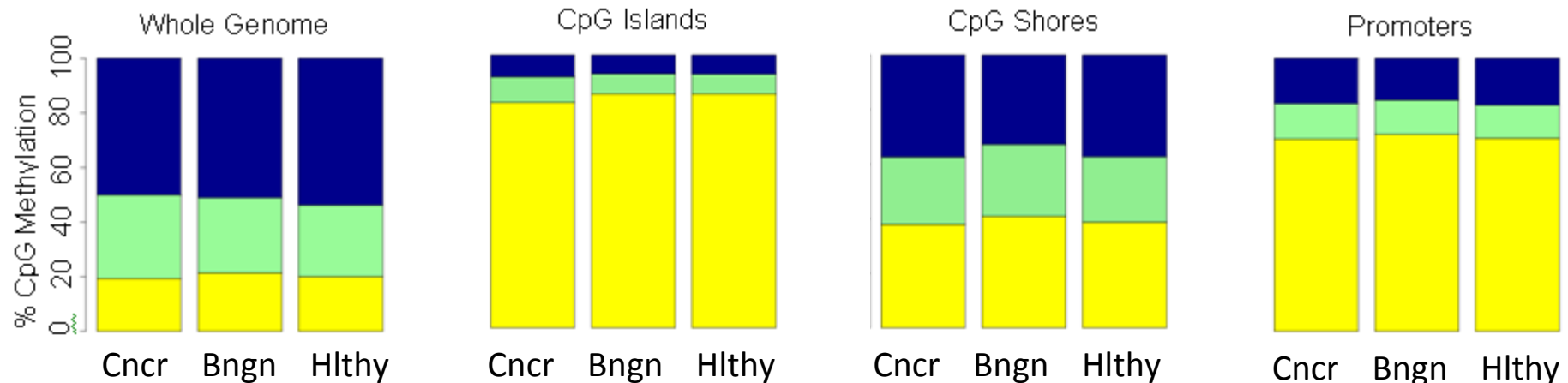


- Aim:
  - Define the methylome associated with a malignant phenotype
  - MeDIP-Seq to identify tumour specific differential methylation
  - Correlation with clinical end points
- Samples:
  - Pools of ten cases per sample cohort
    - Malignant Peripheral Nerve Sheath Tumors (MPNST) – ***Cancer***
    - Benign plexiform neurofibromas – ***Benign***
    - Normal Schwann Cell – ***Healthy***

# Global changes in methylation

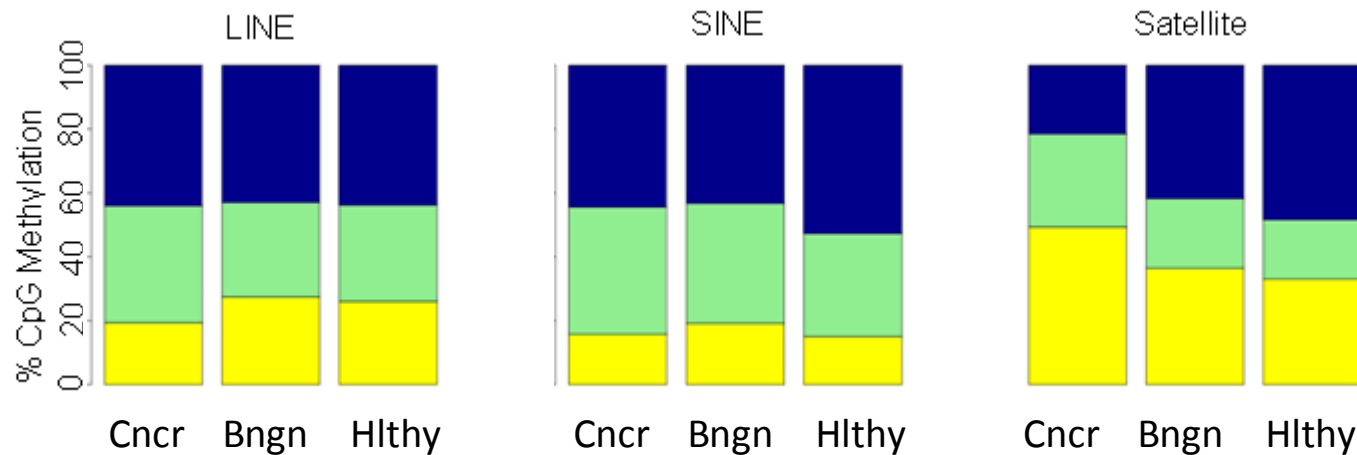


- Methylation state of each CpG site binned
  - Low (<40%), intermediate (40-60%) & high (>60%)
- Studies of other tumour types
  - hallmark of a cancer genome = global hypomethylation

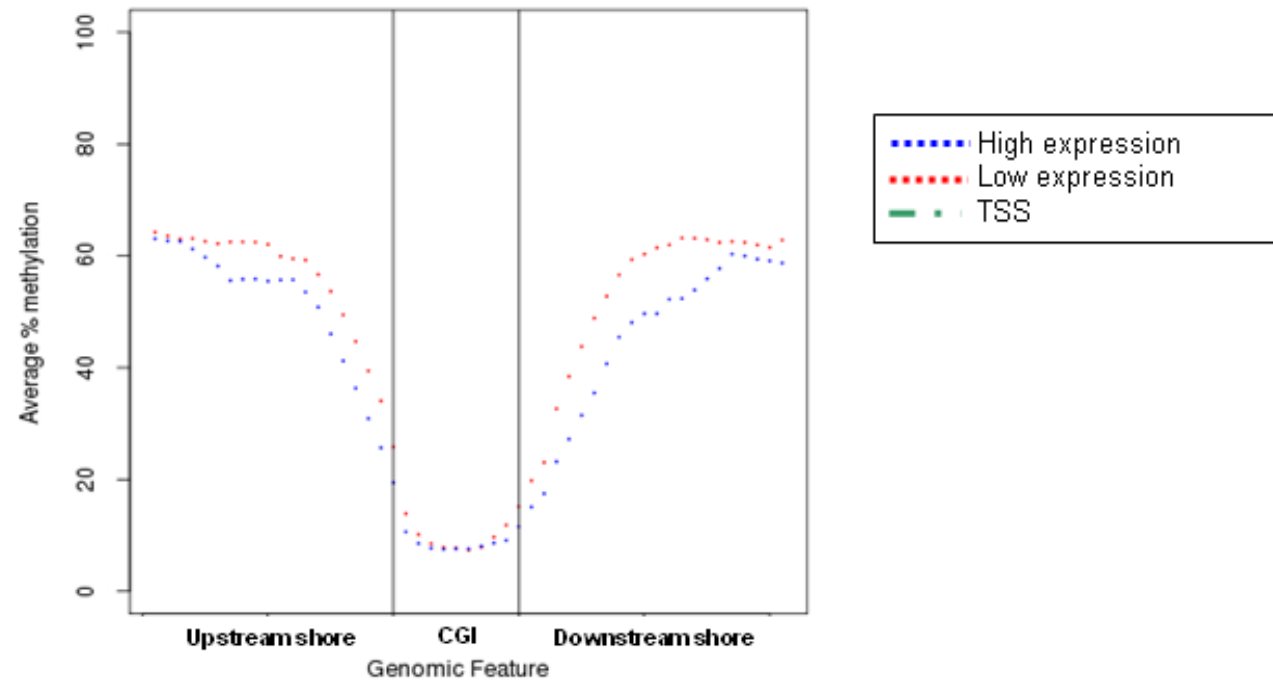


# Global repeat methylation

- Methylation over LINE and SINE repeats change slightly
- Interestingly, LINE repeats appear to lose hypomethylated CpGs
- Satellite repeats show the largest change in global methylation



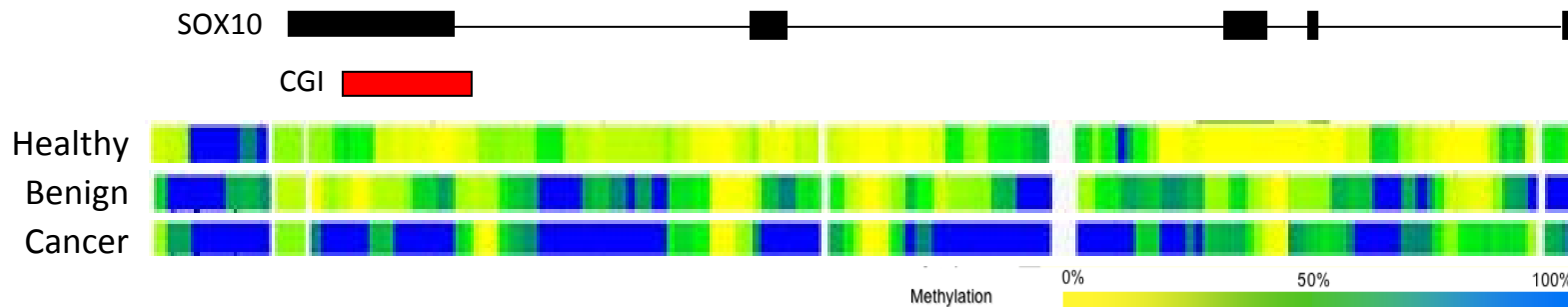
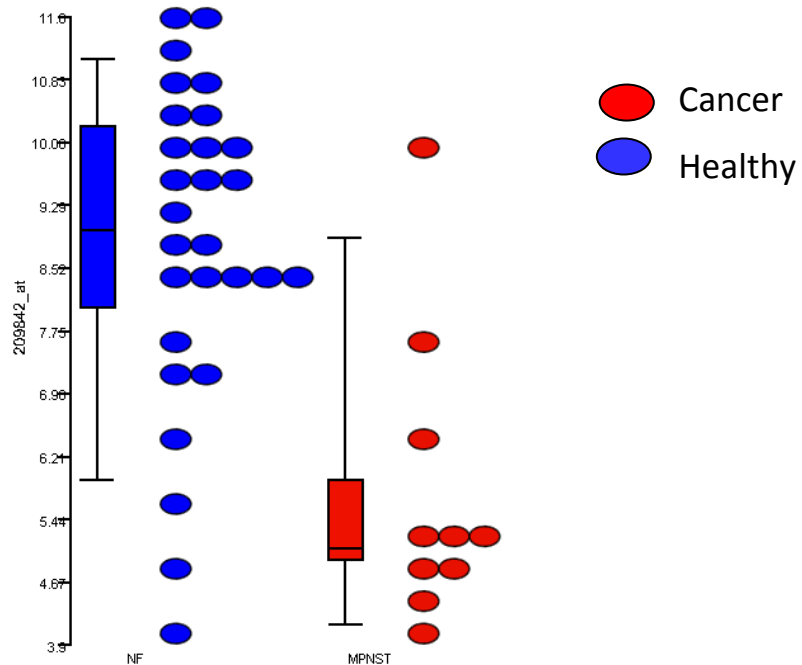
# Effect of methylation at CGIs and CGI shores on gene expression





# SOX10

Feber et al (2010) Submitted



# Acknowledgements



## Lab:



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Li Ning  
Qiuqing Zhang  
Jun Wang

## Babraham Institute, UK:

Steffi Seisenberger

# Questions?



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<http://www.ucl.ac.uk/cancer/research-groups/medical-genomics/index.htm>